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Phenotypic and Molecular Diagnosis of Some Bacterial Species Isolated from Gingivitis from Patients in Al-Rifai District

Noor Abdul Ridha Al-Buhamrah¹, Ebtehal Edrees Shubbar², Widad Sameer jaaz³, Ayat Qasim Owaid⁴

Abstract

Gingivitis is an inflammatory, non-destructive disease mainly caused by microbial plaques. The study was proposed to explore bacterial agents causing gingivitis. Subjects were 100 patients suffering from periodontal infections as well as other dental problems who visited governmental health centers and outpatient clinics in Al-Rifai District during the period from September 2022 to March 2023. Initial characterization of the gingiva was done by examination to exclude periodontitis cases. Those diagnosed as gingivitis patients were 48 patients, comprised of 33 males and 15 females. Gingival specimens was done to proceed with identification, which was performed by cultural, microscopic, utilization of the Vitek compact system, and molecular characterization. 41 specimens gave positive culture on at least one of the used culture media, while 7 specimens showed no growth on any of those media. The positive growth cultures were either pure cultures (28) or mixed cultures (13), which were excluded from the study. The 28 pure cultures were submitted to the vitek-2 compact system. Relying on the latter, identification results showed a high frequency of species belonging to the Gram-negative bacilli. Where the highest percentage was of Enterobacter cloacae (six isolates) (21.43%%), the lowest percentage was one isolate (3.55%) for each of: Aeromonas veronii, Pantoea spp., Laclercia adecarboxylate, Klebsiella pneumonia, Acinetobacter lowifii, Pseudomonas species, Burkholderia gladioli, and Staphylococcus aureus as gram-positive bacteria, and six specimens (21.43%) were not identified. According to age groups, the highest number of infections was in the age group (41-60) with 20 infections. According to sex, the percentage of infections among males was higher 33 (68.75%) than females 15(31.25%). The results of the present study showed an obvious variation in bacterial etiogens that were responsible for the destruction of gingiva among Al-Rifai District patients.

Keywords: Phenotypic, Molecule, Bacterial Species.

Introduction

Gingivitis represents the most prevalent disease of the periodontium and is usually known to be a site-specific inflammatory disease resulting from the buildup of dental biofilm (Zhang et al., 2021). Occasionally, some of the bacteria found in the mouth from gingivitis can easily drip into the bloodstream, causing organ damage. The disease of periodontium may be featured as gingivitis and periodontitis (<u>Murakami et al., 2018</u>). Although gingivitis

¹ Department of Pathological Analyses, Faculty of Science, University of Kufa, Najaf, Iraq, noora.albohamra@uokufa.edu.iq

² Department of Pathological Analyses, Faculty of Science, University of Kufa, Najaf, Iraq

³ Department of Microbiology, College of Medicine, University of Karbala

⁴ Department of Microbiology, College of Medicine, University of Karbala

always precedes periodontitis, the former does not always develop into periodontitis. Gingivitis results from the exacerbation of an inflammatory response against the activity of the oral microbiome. Symptoms are various and range from bleeding and redness to systemic inflammation passing through loss of tooth (Batchelor, 2014; Onyido et al., 2011).

Plaque starts when changes occur in the oral microbiome. When turns from healthy gum to plaque formation, examination of plaque showed little variation of microbes represented by gram-positive rods and cocci. Later on, the community becomes more complex and is dominated by gram-negative rods, spirilli, and spirochetes (Kistler et al., 2013).

Several factors are responsible for the pathogenesis of gingivitis, including microbial plaques, drugs, hormonal fluctuations, malnutrition, and some systemic diseases. Other factors still control the susceptibility and regulatory capacity of microbial pathogens within plaques, like genetic variation as well as epigenetic programs (Wade, 2013).

Various methods have been utilized to determine the microbial pathogens of gingivitis, including culture-based methods and culture-independent molecular methods, which supplied highly accurate results reflecting the huge phylotype variation within the oral microbial community based on 16S rRNA sequencing (Mythri et al., 2015; <u>Wade, 2013</u>).

Good oral hygiene could reverse gingivitis. Treatment is necessary to restore the normal condition of the gingiva and prevent it from developing into periodontitis. The latter is one of the leading causes of tooth loss. (<u>Adams</u> et al., 2000).

Genomic Methods

1- DNA extraction

Genomic DNA was extracted from six bacterial isolates that were not diagnosed with the Vitek 2 system in order to diagnose them by PCR using the boiling method (Dashti et al.,2009).

2- Prepare the polymerase chain reaction mixture

2 microliters of the DNA template were mixed with 10 μ l Master mix ,1 μ l Forward primer , 1 μ l reverse primer and 6 μ l double-distilled water .The final volume was reduced to 20 μ l (Srivastava et al.,2008).

3- PCR amplification

The PCR technique was performed by a gradient thermal cycler. The universal primers Forward 5'- AGAGTTTGATCCTGGCTCAG -3' and reverse primer 5'-CTTGTGCGGGCCCCCGTCAATTC-3') of primer targeted 16S rRNA. Amplification was performed by initial denaturation at (94°C for 3 min), followed by (30 cycles) of denaturation at (94°C for 30 sec), annealing temperature was(55°C for 30 sec) and extension at(72°C for 1 min). Final extension was at(72°C for 10 min) (Dos Santos et al.,2018; Srivastava et al.,2008).

Agarose gel electrophoresis

The agarose gel electrophoresis technique was carried out for the detection of amplicon, as mentioned by (Sambrook and Russell 2001).

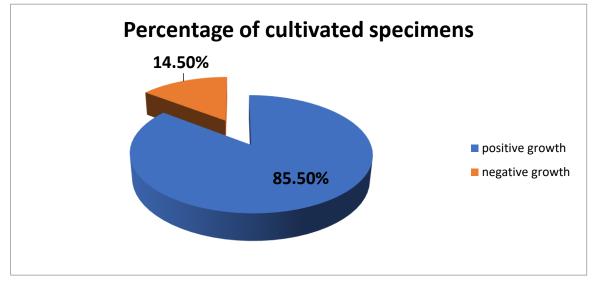
DNA sequencing of the 16S rDNA fragment

DNA samples of the 16S rDNA gene with primers (forward and reverse) were sent to Macrogen Inc (South Korea Geumchen, Seoul) and the results were then analyzed.

RESULTS AND DISSCUSSION

Isolation and Identification

In this study, forty-eight swabs have been collected from forty-eight patients diagnosed with gingivitis. Forty-one (85.5%) specimens showed positive growth on different culture, while seven (14.5%) specimens showed negative growth on different culture. (Figure -1) shows the percentage of Gingivitis specimens growth cultures



(Figure -1) shows the percentage of Gingivitis specimens growth cultures

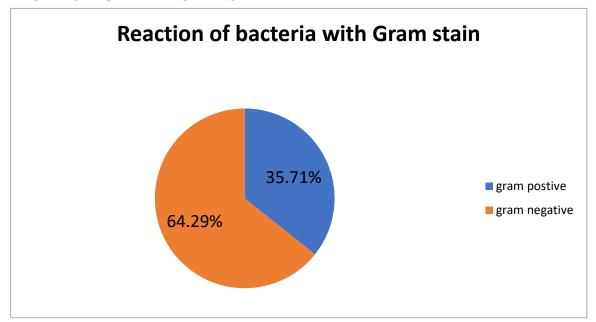
Jaseem and Yasin (2023) found (75%) specimens showed positive bacterial growth cultures, while (25%) specimens showed no bacterial growth cultures.

Cultures varied from pure culture, characterized by the appearance of a single, defined type of colony, reflecting their return to a single type of microorganism. The total number of such cultures was 28 (68.29%); to those that showed multiple types of colonial appearance, which means that there are multiple microorganisms forming them, represented 13 (31.70%). The latter were excluded (Table -1). Studies showed that approximately half of oral microbes cannot be cultivated or have not been cultivated in laboratory conditions (Wade, 2004). Multiple bacterial growths support the concept of the complexity of the oral cavity microbial world. In a study by Aas et al.,(2005), results estimated from 34 to more than 70 different phylotypes at the level of species in each single individual included in the study were determined by molecular-dependent methods.

No. of Positive	8	No. of Negative Samples (%))
Samples (%)			
41 (85.5)		7(14.5)	
No. of Pure cultures (%)	No. of mixed cultures isolates (%)		
28 (68.3)	13(31.7)		

(Table -1): The Number and Percentage of Positive and Negative Culture Samples

After that, a microscopic examination of all bacterial isolates was performed using the Gram stain to differentiate between positive and negative bacteria to interact with the Gram stain and to know the shape, size and arrangement of the bacteria. The results showed that 10(35.71%) bacterial isolates were positive for the Gram stain and that 18 (64.29\%)



bacterial isolates were negative for the Gram stain. (Figure -2) shows the percentage of Gingivitis gram positive and gram negative.

(Figure -2) shows the percentage of Gingivitis gram positive and gram negative

a broad spectrum of bacteria including, gram-negative bacilli were found as the predominant microbial populations in the gingival specimens. The differences observed between these results can be addressed to the area of sampling on the gingival plaques. The result obtained by this study is consistent with the results of (Azadeh et al., 2011) while don't consistent with those reported by (Leo ,1981) and (Moening, 1989) who emphasized the importance of gram positive cocci in the initiation of dental plaque formation, leading to gingivitis.

After that, the diagnosis was made using the Vitek 2 compact system to accurately diagnose all bacterial isolates .The results revealed that 10 (35.71%) of bacteria isolates which were gram positive bacteria Staphylococcus aureus 1 (3.57%), Un-identified. While gram negative were found in 18 (64.29%) isolates , represented by 6 (21.44%) of Enterobacter cloacae spp, 3 (10.72%) isolates of Klebsiella pneumonia, 2 (7.14%) Acinetobacter lowifii , Pseudomonas species and Burkholderia gladioli, 1 (3.57%) isolates of Aeromonas veronii ,Pantoea spp and Laclercia adecarboxylate respectively. (Table -2) exhibit types and numbers of bacterial isolates identified in the study.

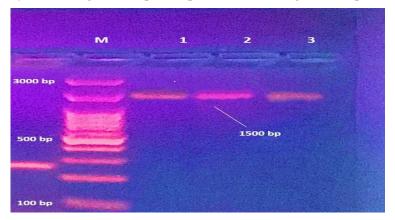
No	Bacterial isolates	No (%)
1	Enterobacter cloacae spp	6 (21.44)
2	Klebsiella pneumonia	3 (10.72)
3	Acinetobacter lowifii	2 (7.14)
4	Pseudomonas species	2 (7.14)
5	Burkholderia gladioli	2 (7.14)
6	Aeromonas veronii	1 (3.57)
7	Pantoea spp	1 (3.57)
8	Laclercia adecarboxylate	1 (3.57)

(Table-2): Types and Numbers of Bacterial Isolates Collected in This Study.

9	Staphylococcus aureus	1 (3.57)
10	Un-identified	9 (32.14)
	Total	28(100)

This study is in agreement with Alghamdi, (2022) who showed in isolation of species of different families including six families of the gram positive bacteria; Leuconostocaceae, Listeriaceae, Streptococcaceae, Staphylococcaceae, Bacillaceae, Corynebacteriaceae, and 5 families of gram negative; Neisseriaceae, Enterobacteriaceae, Pseudomonadaceae, Yersiniaceae, and Moraxellaceae,. The overall dominated family observed was Enterobacteriaceae (19.36%) having Escherichia coli as the most prevalent specie (53.06%) followed by Klebsiella pneumoniae (28.57%). Al-Shammarie and Maaroof (2020) Perkowski et al., (2019) also found many different bacterial species that cause gingivitis.

After that Molecular identification of those which could not be identified by vitek-2 system. Where there were 9 isolates, the vitek -2 system gave un -identified results for them. Those were submitted to molecular identification by 16S rRNA . only three gave positive result by PCR that gave the specific product size after gel electrophoresis. (Figure-1).



(Figure -3): Gel Electrophoresis of 16SrRNA Sequence result after PCR

DNA Sequencing results showed that two of them were S. aureus and on was S.equoeumas shown in figure (4), (5) and (6). This method is one of the most important ways to identify the identity of pathogenic bacteria. This method also detects the occurrence of genetic mutations in resistance genes and other genes, by identifying the sequence of the nitrogenous bases of the gene. Thus, the number of diagnosed bacterial isolates in this study became 22 (78.57%) bacterial isolates out of 28 bacterial isolates, while the number of undiagnosed isolates was 6 (21.43%) bacterial isolates.

Dow	nload	✓ GenBank Gr	aphics			
Staph	yloc	occus aureus sti	rain L10EW2 16S	ribosomal RNA	gene, partial s	equence
Sequer	nce ID:	MW295826.1 Len	gth: 637 Number of	Matches: 1		
Range	1: 224	to 265 GenBank G	iraphics		▼ <u>Next Match</u>	A Previous Match
Range Score	1: 224	to 265 <u>GenBank</u> <u>G</u> Expect	Identities	Gaps	▼ <u>Next Match</u> Strand	Previous Match
		Expect		Gaps 1/42(2%)		Previous Match
Score		Expect 1e-07	Identities	1/42(2%)	Strand	Previous Match

(Figure -4) Sequence analysis of the 16SrRNA gene of bacteria S. aureus in isolation number 1

Range	1: 466	to 1396 GenE	lank	Graphics		w Next I	Match & Previous Ma
Score 1541 b	oits(834) Expe 0.0	ect	Identities 913/948(96%)	Gaps 18/948(1%)	Strand Plus/Minu	15
Query	36	ACTCTCGCCGG	CTTO	GGGAGTTACAAACTCT	CGTGGTGTGACGGGCGGT	GTGTACAAGAC	95
Sbjct	1396	ACTC-CACCGO	CTTO	GGGTGTTACAAACTCT	GTGGTGTGTGACGGGCGGT	GTGTACAAGAC	1338
Query	96	CCGGGAACGTA	TTC	CCGTAGCATGCTGATC	TACGATTACTAGCGATTO	CAGCTTCATGT	155
Sbjet	1337	CCGGGAACGTA	TTC	CCGTAGCATGCTGATC	TACGATTACTAGCGATTO	CAGCTTCATGT	1278
Query	156	AGTCGAGTTGC	AGAG	TACAATCCGAACTGAG	ACAACTTTATGGGATTI	GCATGACCTCG	215
Sbjet	1277	AGTCGAGTTGC	AGAG	TACAATCCGAACTGAG	ACAACTTTATGGGATTT	GCATGACCTCG	1218
Query	216				AGCACGTGTGTAGCCCA4		275
Sbjet	1217				AGCACGTGTGTAGCCCAA		1158
Query	276	CATGATGATTI	IGACO	TCATCCCCACCTTCCT	CONTINUES	TCAACCTAGAG	335
Sbjct	1157	CATGATGATT	GACO	TCATCCCCACCTTCCT	CGGTTTGTCACCGGCAG	TCAACCTAGAG	1098
Query	336	TGCCCAACTAA	ATG	TGGCAACTAAGTTTAA	SGGTTGCGCTCGTTGCGG	GACTTAACCCA	395
Sbjct	1097	TGCCCAACTAA	ATG	TGGCAACTAAGTTTAA	GGTTGCGCTCGTTGCGG	GACTTAACCCA	1038
Query	396				SCACCACCTGTCACTTTC		455
Sbjet	1037	ACATCTCACGA	CACO	GAGETGACGACAACCAT	GCACCACCTGTCACTTTG	TCCCCCGAAGG	978
Query	456				TGTCAAGATTTGGTAGGG		515
Sbjct	977	GGAAGGCTCTA	TCTO	TAGAGTTTTCAAAGGA	TGTCAAGATTTGGTAAGO	TTCTTCGCGTT	918
Query	516				STGCGGGCCCCCGTCAAT		575
Sbjct	917	GCTTCGAATTA	AACO	ACATGCTCCACCGCTT	GTGCGGGTCCCCGTCAAT	TCCTTTGAGTT	858
Juery	576	TCAACCTTGCO	GTC	GAACTCCCCAGGCGGAG	TGCTTAATGCGTTAGCTG	GCAGCACTAAGG	635
Sbjet	857	TCAACCTTGCC	GTCC	TACTCCCCAGGCGGAG	I GCTTAATGCGTTAGCTC	CAGCACTAAGG	798
Juery	636				STTTACGGCGTGAACTAG		695
Sbjct	797	GGCGGAAACCO	Let.	ACACTTAGCACTCATO	STTTACGGCGTGGACTAC	CAGGGTATCTA	738
Juery	696				AGCGTCAGTTACAAACCA	GAAAATCCCCC	755
bjet	737	ATCCTGTTTGA	HLL	CACGCTTTCGCACATC	AGCGTCAGTTACAGACCA	GAAAGTCGCC-	679
Juery	756				GCATTTCACCGCTACACA		815
Sbjct	678	TTCGCCACTGO	TGTT	TCCTCCA-TATCTCTGC	SCATTTCACCGCTACACA	TGGAATTCCAC	628
Juery	816	TTTTCCTCTTC	TGTA	ACTCAAGTTTCCCAGTT	TCCAATGACCCTCCACGO	TTGAGCCGTGG	875
Sbjct		111 1111111	1111		TCCAATGACCCTCCACGG		561
Juery					AACGC-CGCCTTTTACCO		934
Sbjct	560	THEFT FEE			A-CGCGCGC-TTT-ACGC	1111 11111	508
Query						982	1707038
	507			-GCCACC-TACGT-ATT		466	

(Figure -5) Sequence analysis of the 16SrRNA gene of bacteria S.equoeum in isolation number 2

100	viloco			ribosomal RNA o	ene nar	tial sequence	Next <u>Next</u> <u>Previous</u>	- Descriptions
			ogth: 1454 Number of		ferre, pur			
Range	1: 330	to 1436 GenBank	Graphics		V Next	Match A Previous Match		
Score 1978 b	bits(107	Expect (1) 0.0	Identities 1100/1113(99%)	Gaps 6/1113(0%)	Strand Plus/Min	ius		
Query				TCATACCGGCTTCGGGTG				
	1436	CCACTTCGACGGCT	AGC-TC-CTAAAAGGTTA	TC-CACCGGCTTCGGGTG	TTACAAACT			
uery	73 1379	111111111111111	1111111111111111111111	CGGGAACGTATTCACCGTA				
uery				STCGAGTTGCAGACTACAA		1920		
	1319			TEGAGTTGCAGACTACAA				
Juery				GTTTCGCTGCCCTTTGTA		252		
	1259	AGAACAACTTTATG	GGATTTGCTTGACCTCGC	GTTTCGCTGCCCTTTGTA	HIGTCCATT	1200		
Query		GTAGCACGTGTGTA	GCCCAAATCATAAGGGGGCA	ATGATGATTTGACGTCATC	CCCACCTTC	312		
	1199	GTAGCACGTGTGTA	GCCCAAATCATAAGGGGGCA	TGATGATTTGACGTCATC	CCCACCTTC	1140		
Query	313			SCCCAACTTAATGATGGCA		372		
Sbjct	1139	CTCCGGTTTGTCAC	CGGCAGTCAACTTAGAGT	SCCCAACTTAATGATGGCA.	ACTAAGCTT	1080		
Query	373	AAGGGTTGCGCTCG	TTGCGGGACTTAACCCAA	ATCTCACGACACGAGCTG	ACGACAACC	432		
Sbjct	1079	AAGGGTTGCGCTCG	TTGCGGGACTTAACCCAA	CATCTCACGACACGAGCTG	ACGACAACC	1020		
Query	433	ATGCACCACCTGTC	ACTITGTCCCCCGAAGGGG	GAAAGCTCTATCTCTAGAG	TTGTCAAAG	492		
Sbjet	1019	ATGCACCACCTGTC	ACTTTGTCCCCCGAAGGG	GAAAGCTCTATCTCTAGAG	TTGTCAAAG	960		
Query	493	GATGTCAAGATTTG	GTAAGGTTCTTCGCGTTG	TTCGAATTAAACCACATG	TCCACCGC	552		
Sbjct	959	GATGTCAAGATTTG	GTAAGGTTCTTCGCGTTG		CTCCACCGC	966		
Query	553		GTCAATTCCTTTGAGTTT	AACCTTGCGGTCGTACTC	CCCAGGCGG	612		
Sbjct	899	ttätäcäääteee	GTCAATTCCTTTGAGTTT	AACCTTGCGGTCGTACTC	CCAGGCGG	840		
Query	613			GCGGAAACCCCCTAACACT		672		
Sbjct	839	AGTGCTTAATGCGT	tágétőékésékétákégés	GGGAAACCCCCTAACACT	TAGCACTCA	780		
Query		1111111111111111		CCTGTTTGATCCCCACGC	1111111111	732		
Sbjct	779	TCGTTTACGGCGTG	GACTACCAGGGTATCTAAT	CCTGTTTGATCCCCACGC	TTTCGCACA	728		
Query				GCCACTGGTGTTCCTCCA				
sbjct				GCCACTGGTGTTCCTCCA				
Query				CTCTTCTGCACTCAAGTT				
Sbjct				CTCTTCTGCACTCAAGTT				
Query Sbjct		1111111111111111				540		
Juery				TTGCCACCTACGTATTACC				
sbjct				TTGCCACCTACGTATTACC				
Query				FACCGTCAAGATGTGCACA				
Sbjct		TELEVISION	TITLETTITLETT	TACCGTCAAGATGTGCACA				
	1033							
Sbjct		ACATATGTTCTTCC	CTAA-TAACAGAGTTTTA	CGATCCGAAGACCTTCATC	ACTCACGCG	361		
	1093	GGGTTGCTCCGTCA	GGCTTTTCGCCCATTTGC	5 1125				
Sbjct		1 1111111111111	GGCTTT-CGCCCATT-GC					

(Figure -4) Sequence analysis of the 16SrRNA gene of bacteria S. aureus in isolation number 3 $\,$

In a study by Al-Abdul et al., (2017) performed in AL-Basrah showed that Staphylococcus spp. Were the most commonest isolates. Similarly in another study in Kufa study S. aureus was the commonest in acute gingivitis.

The objective of this study was to determine the number and frequencies of bacterial populations associated in dental plaques among Al-Rifai District patients with gingivitis. Some other studies have also been conducted to profile the pathogenic bacteria isolated from gingivitis plaques among different groups of populations in different cities of Iraq (Ibrahim and Hussien, 2018; Jabuk et al.,2015; Najm and Younis,2009). However, in current study,

Distribution of Patients According Age and Sex

According to age groups, the highest number of infections was in the age group (41-60) where it was 20 infections, while the other age groups had the same infections number 14 infections each (Table-3).

In general the results of this study have been conclude that the infection increases with the advanced stages of age , and this is consistent with many studies (Ababneh et al.,2012; Stamm, 1986).

(Table -5). Distribution of Oligival 1 attents /	According to Age
Age group	No. of Patients
1 years -20	14
21-40	14
41-60	20
Total	48

(Table -3): Distribution of Gingival Patients According to Age

As for infections distribution among males and females, the percentage of infections in males was higher 33(68.75) % males than 15(31.25) % females as shown in (Table- 4). At different age stages there an obvious lower frequency of gingivitis in females as compared with males despite the fact that females are influenced by hormonal changes. This may be attributed better teeth health care as well as the presence of physiologic difference between the sexes (Newman et al ., 2006). More recent native study revealed that males showed higher gingival index than females with no significant differences which may be due to the population samples were taken from Dentistry College (Wais et al ., 2023). It was also the same in a study in Sullaimaniya (Hamid et al ., 2020; Kinane et al ., 2017 Humphrey et al ., 2008)

(Table -4) Distribution of Gingival Patients Ac

Sex	Number of Patients (%)
Male	33 (68.75)
Female	15 (31.25)
Total	48 (100%)

Conclusions

The results of present study indicate that a wide range of communicable pathogenic bacteria are responsible for gingivitis and its progression among residents of Al-Rifai District. The oral microbiome is a complex community which could be used further to monitor health status.

References

Aas, J. A., Paster, B. J., Stokes, L. N., Olsen, I., & Dewhirst, F. E. (2005). Defining the normal bacterial flora of the oral cavity. Journal of clinical microbiology, 43(11), 5721-5732.

- Ababneh, Khansa ; Abu Hwaij, Zafer & Khader, Yousef.(2012). Prevalence and risk indicators of gingivitis and periodontitis in a multi-centre study in North Jordan: a cross sectional study. BMC oral health, 12(1): 1-8.
- Abbas, M. H., Al-Yasseen, A. K., & Alhamadi, W. W. (2017). Distribution of Granulicatella adiacens and Porphyromonas gingivalis among ortho and non-orthodontic Patients with Gingivitis in Kufa City/Iraq. Al-Qadisiah Medical Journal, 13(23).
- Adams, D.; Barrington, E., & Caton, J. (2000). col. Parameter on Plaque-Induced gingivitis. J Periodontol, 71(5): 851-852.
- Al-Abdul, A. A., & Hussein, I. K. (2017). Isolation and Identification of Bacteria from Diabetic and Non-Diabetic Patients With Periodontitis. Donish J. of Microbiol. And Biotech. Res, 4(2), 4-9.
- Alghamdi, S. (2022). Isolation and identification of the oral bacteria and their characterization for bacteriocin production in the oral cavity. Saudi Journal of Biological Sciences, 29(1), 318-323.
- Al-Shammarie, Z. Q., & Maaroof, M. N. (2020). Isolation and identification of some bacterial species causing gingivitis in women over the age of 45 and molecular detection of its virulence factors. Journal of Education and Scientific Studies, 4(15).
- Azadeh, M., KASRA, K. R., Naghavi, N. S., Ghalayani, P., & Salamat, F. (2011). the profile of pathogenic bacteria isolated from dental plaqueinduced gingivitis.
- Batchelor, P. (2014). Is periodontal disease a public health problem?. British dental journal, 217(8), 405-409.
- Dashti, A. A., Jadaon, M. M., Abdulsamad, A. M., & Dashti, H. M. (2009). Heat treatment of bacteria: a simple method of DNA extraction for molecular techniques. Kuwait Med J, 41(2), 117-122.
- Dos Santos, H. R. M., Argolo, C. S., Argôlo-Filho, R. C., & Loguercio, L. L. (2019). A 16S rDNA PCR-based theoretical to actual delta approach on culturable mock communities revealed severe losses of diversity information. BMC microbiology, 19(1), 1-14.
- Hamid, R., Mudher, S., and Ali, S.(2020). Caries Index, Root Caries Index and Gingival Index in Immigrants at the Camp of Arbat in Sulaimaniya Governorate, Iraq. International Medical Journal. 27 (3): 357 - 359.
- Humphrey, L. L., Fu, R., Buckley, D. I., Freeman, M., and Helfand, M. (2008). Periodontal disease and coronary heart disease incidence: a systematic review and meta-analysis. J. Gen. Intern. Med. 23, 2079–2086. doi: 10.1007/s11606-008-0787-6.
- Ibrahim, Jihan A. & Hussien, Baydaa .(2018). Gingival Condition and Enamel Defect Among Secondary School Female Students in Kirkuk City/Iraq. Journal of baghdad college of dentistry, 30 (1): 80-85.
- Jabuk, Sura; Hussien ,Rafla'a ; ,Zahraa , Altaee ; Hawraa and Najam, Noor.(2015). Isolation and identification of bacteria and parasite from teeth caries and periodontal. Advances in Environmental Biology, 9(22): 50-53.
- Jaseem, R. M., & Yasin, L. Q. (2023). Isolation and identification of some predominant bacteria and assessment of TNF-α level in serum of patients with gingivitis. Tikrit Journal of Pure Science, 28(5).
- Jasim, R. A., & Jasim, R. M. (2020). Isolation and identification of pathogenic bacteria infection and treatment by active substances isolated from agaricus bisporus fungi. Plant Archives, 20(1), 2732-2735.
- Kinane, D. F., Stathopoulou, P. G., and Papapanou, P. N. (2017). Periodontal diseases. Nat. Rev. Dis. Primers 3:17038
- Kistler JO, Booth V, Bradshaw DJ, Wade WG (2013) Bacterial Community Development in Experimental Gingivitis. PLOS ONE 8(8): e71227.Microbiol 54: 93–106.
- Löe, H. (1981). The role of bacteria in periodontal diseases. Bulletin of the World Health Organization, 59(6), 821.

- Moening, J. E., Nelson, C., & Kohler, R. (1989). The microbiology and chemotherapy of odontogenic infection. J. Oral Maxillofac. Surg, 47, 976-985.
- Murakami, S., Mealey, B. L., Mariotti, A., and Chapple, I. L. C. (2018). Dental plaque-induced gingival conditions. J. Periodontol. 89(Suppl. 1), S17–S27.
- Mythri, Sarpangala ;Arunkumar, Suryanarayan; Hegde, Shashikanth ; Rajesh Shanker; Munaz, Mohamed & Ashwin, Devasya.(2015). Etiology and occurrence of gingival recession-An epidemiological study. Journal of Indian Society of Periodontology, 19(6): 671.
- Najm, Mohanad & Younis, Wasan.(2009). The prevalence of oral and dental developmental anomalies among 14-17 years Iraqi students in Missan governorate. J Bagh Coll Dentistry , 21(4):90-5.
- Newman,M., Takei, H., Carranza, F., et al.(2006) Clinical periodontology, ed 10, Philadelphia, , WB Saunders
- Oh, T. J., Eber, R., & Wang, H. L. (2002). Periodontal diseases in the child and adolescent. Journal of clinical periodontology, 29(5), 400-410.
- Onyido, Angus; Amadi, E.; Olofin, I.; Onwumma, A.; Okoh, I., & Chikwendu, C. (2011). Prevalence of Entamoeba gingivalis and Trichomonas tenax among dental patients attending Federal School of Dental Technology and Therapy clinic, Enugu, Nigeria. Oral diseases, 11(49.2): 35-0.
- Perkowski, K., Balraza, W., Conn, D. B., Marczyńska-Stolarek, M., & Chomicz, L. (2019). Examination of oral biofilm microbiota in patients using fixed orthodontic appliances in order to prevent risk factors for health complications. Annals of Agricultural and Environmental Medicine, 26(2), 231-235.
- Sambrook , J. and Russell , D. (2001) .Molecular Claning Laboratory Manual . 3rded . Cold Spring Harbor , New York . USA . 1044.
- Srivastava, S., Singh, V., Kumar, V., Verma, P. C., Srivastava, R., Basu, V., ... & Rawat, A. K. (2008). Identification of regulatory elements in 16S rRNA gene of Acinetobacter species isolated from water sample. Bioinformation, 3(4), 173.
- Stamm, J. W. (1986). Epidemiology of gingivitis. J. Clin. Periodontol. 13, 360–366. doi: 10.1111/j.1600-051x.1986.tb01473.
- Tringe, Susannah ; Zhang, Tao ; Liu, Xuguo; Yu, Yiting; Lee, Wah Heng ; Yap, Jennifer and Suan, Sim Tiow. (2008). The airborne metagenome in an indoor urban environment. PloS one, 3(4): e1862.
- Wade, W. G. (2004). Non-culturable bacteria in complex commensal populations. Advances in applied microbiology, 54(54), 93-106.
- Wade, W. G. (2013). The oral microbiome in health and disease. Pharmacological research, 69(1), 137-143.
- Wais, Z., Salman, O., Khafaji, S., Wais, A. (2023). Gingivitis and enamel defect among students of Dentistry College in Babylon. Journal of Pakistan Association of Dermatologists.;33(4):1330-1334.
- World Health Organization. (2003). Manual for the laboratory identification and antimicrobial susceptibility testing of bacterial pathogens of public health importance in the developing world: Haemophilus influenzae, Neisseria meningitidis, Streptococcus pneumoniae, Neisseria gonorrhoea, Salmonella serotype Typhi, Shigella, and Vibrio cholerae (No. WHO/CDS/CSR/RMD/2003.6). World Health Organization.
- Zhang, J., Sun, M., Zhao, Y., Geng, G., & Hu, Y. (2021). Identification of Gingivitis-Related Genes Across Human Tissues Based on the Summary Mendelian Randomization. Frontiers in Cell and Developmental .Biology, 8, 624766.
- Zhang, Z. M., Tan, J. X., Wang, F., Dao, F. Y., Zhang, Z. Y., and Lin, H. (2020). Early diagnosis of hepatocellular carcinoma using machine learning method. Front. Bioeng. Biotechnol. 8:254. doi: 10.3389/fbioe.2020.00254.